

APIENE ESTERS FROM *FERULA HAUSSKNECHTII*

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Key Word Index—*Ferula haussknechtii*; Apiaceae; sesquiterpene esters; apienes.

Abstract—Ten new apiene-type humulenoid esters were isolated from the root of *Ferula haussknechtii*. The structures were elucidated by spectroscopic methods.

INTRODUCTION

In continuation of our investigation of *Ferula* species which are indigenous to the Anatolia region of Turkey [1-4], we investigated *F. haussknechtii* Wolff. ex Rech. *Ferula* species are known to be a rich source of different classes of sesquiterpenes such as daucanes, germacrane, eudasmanes, guiananes, elemenes, humulenes and himacalenes [5]. Although the occurrence of humulenes is relatively rare in the genus *Ferula*, α -humulenes from *F. juniperina* [6-8], *F. xeromorpha* [9, 10] and *F. tschatalensis* [11], as well as γ -humulenes from *F. ceratophylla* [12], have been reported previously.

RESULTS AND DISCUSSION

The dichloromethane extract of the root of *F. haussknechtii* afforded ten new sesquiterpene esters in small amounts; their structures were elucidated by spectral techniques, especially stereochemical data obtained with the relatively new techniques such as 2D NMR homo- and heteronuclear COSY and high resolution NOESY experiments.

The IR spectra of the benzoate (1) ($C_{22}H_{28}O_2$), *p*-hydroxybenzoate (2) ($C_{22}H_{28}O_3$) and vanillate (3) ($C_{23}H_{30}O_4$) derivatives of servanol all exhibited absorptions for an aromatic acyl group. The 1H NMR (Table 1), ^{13}C NMR (Table 2) and MS data for 1, 2 and 3 confirmed the presence of benzoate, *p*-hydroxybenzoate and vanillate acyl groups, respectively, as well as the same sesquiterpene nucleus for all three. A total of four degrees of unsaturation were assigned for this nucleus based on the ^{13}C NMR and mass spectral data. The 1H NMR and ^{13}C NMR spectra of 1-3 indicated the presence of an exocyclic methylene group (δ 4.96, 4.90, each 1H, *d* and 148.3, *s*; 114.7, *t*), a trisubstituted double bond (δ 5.32, 1H, *t*; 1.48, 3H, *d* and 126.9, *d*; 135.5, *s*) and a *trans* disubstituted double bond (δ 6.04 and 5.59, each 1H, *d*, *J* = 16.1 Hz; 138.5, *d* and 128.3, *d*), as well as two geminal methyl groups (δ 1.16 and 1.07, each 3H, *s*; both at 26.0, *q*). The presence of these three double bonds indicated a monocyclic structure for all three compounds; moreover, the presence of two geminal methyl groups instead of an isopropyl group clearly demonstrated the γ -humulenoid-type structure for 1-3. The location of the aromatic acyl

groups and the structural relationship of these three compounds was deduced from 2D NMR homonuclear COSY-45 experiments. Nevertheless, the spectral data did not establish all stereochemical matters. Interestingly, in the 1H NMR spectra of 1-3 the exocyclic methylene proton signals appear as two doublets, whereas the same signals in the spectra of all previously known γ -humulenes except for the compound obtained from *F. ceratophylla*, which probably has the same stereochemical features as 1-3, were reported as broad singlets [13, 14]. The 2D NMR homonuclear COSY-45 experiment confirmed the presence of geminal coupling between the exocyclic methylene protons but no other long range couplings with these protons. This feature is probably indicative of a different conformation of 1-3 relative to previously reported γ -humulenes. Further information regarding the stereochemistry of 1-3 was obtained by a series of NOESY experiments. The 1H NMR NOESY experiments of 3 verified an *s-trans* for the conjugated 4(15),5-diene system and β orientation for the C-7 acyl group.

The IR spectra of the benzoate (4) ($C_{22}H_{28}O_3$) and vanillate (5) ($C_{23}H_{30}O_5$) of kurubasch aldehyde showed an α,β -unsaturated aldehyde (1695 cm^{-1}) and aromatic acyl group absorptions. The aromatic acyl groups of 4 and 5 were assigned as benzoate and vanillate, respectively, on the basis of their 1H , ^{13}C NMR and mass spectral data. Similarity of their spectra, except for the acyl group signals, confirmed the same sesquiterpene skeleton in both 4 and 5. In addition to the other spectral data, an α -humulenoid structure for 4 and 5 was deduced from the 2D NMR homonuclear COSY-45 experiments. In the 1H NMR spectra of 4 and 5 the downfield position of the H-5 signal (δ 6.50, 1H, *d*) and the presence of an aldehyde proton signal (δ 9.4, 1H, *d*) instead of a second vinylic methyl signal expected for an α -humulene, clearly indicated that in 4 and 5 the C-4 methyl of α -humulenes had been replaced with an aldehyde group. Furthermore, the doublet at δ 5.7 (1H), which coupled with the former H-5 signal, at δ 6.50, confirmed the location of the aromatic acyl groups to be C-6 in both compounds. The NOESY experiments of 4 (Table 3) showed that both the $\Delta^{1,10}$ and Δ^4 double bonds had the *E* configuration and that the stereochemistry of the acyl group at C-6 is β .

Kurubaschic acid angelate (6) ($C_{20}H_{30}O_4$) and benzoate (7) ($C_{22}H_{28}O_4$) exhibited similar spectral data to

Table 1. ^1H NMR spectra data of compounds 1–10

| H | 1 (200 MHz) | 2 (200 MHz) | 3 (500 MHz) | 4 (500 MHz) | 5 (200 MHz) |
|------------------|----------------------------|---------------------------|------------------------------|---------------------------------|------------------------------|
| 1 | 5.32 <i>br t</i> (8.5) | 5.31 <i>br t</i> (8.4) | 5.32 <i>br t</i> (8.5) | 5.32 <i>br dd</i> (6.3; 5.8) | 5.24 <i>br t</i> (7.6) |
| 2 | 2.22 <i>m</i> | 2.21 <i>m</i> | 2.22 <i>m</i> | 2.28 <i>m</i> | 2.26 <i>m</i> |
| 2' | 2.33 <i>m</i> | 2.32 <i>m</i> | 2.33 <i>m</i> | 2.19 <i>m</i> | |
| 3 | | | | 2.61 <i>td</i> | |
| 3' | 2.38 <i>m</i> | 2.36 <i>m</i> | 2.38 <i>m</i> | (4.1; 12.6) | 2.60 <i>m</i> |
| | | | | 2.54 <i>dt</i> (3.8; 12.6) | |
| 5 | 6.04 <i>d</i> (16.1) | 6.04 <i>d</i> (16.1) | 6.04 <i>d</i> (16.1) | 6.49 <i>d</i> (10.3) | 6.50 <i>d</i> (10.3) |
| 6 | 5.59 <i>d</i> (16.1) | 5.58 <i>d</i> (16.1) | 5.59 <i>d</i> (16.1) | 5.72 <i>d</i> (10.3) | 5.70 <i>d</i> (10.3) |
| 7 | | | | 1.33 <i>m</i> | |
| | 4.68 <i>br d</i> | 4.68 <i>br d</i> | 4.68 <i>br d</i> | | 1.38 <i>m</i> |
| 7' | (6.9) | (6.8) | (6.9) | 1.20 <i>m</i> | |
| 8 | 2.20 <i>m</i> | 2.19 <i>m</i> | 2.24 <i>m</i> | 1.64 <i>m</i> | 1.60 <i>m</i> |
| 8' | 1.91 <i>m</i> | 1.90 <i>m</i> | 1.92 <i>m</i> | 1.36 <i>m</i> | 1.38 <i>m</i> |
| 9 | 2.17 <i>m</i> | 2.17 <i>m</i> | 2.17 <i>m</i> | 2.20 <i>m</i> | 2.21 <i>m</i> |
| 9' | 1.58 <i>m</i> | 1.58 <i>m</i> | 1.58 <i>m</i> | 1.46 <i>dt</i> (2.1; 13) | 1.48 <i>dt</i> (2.1; 13) |
| 12 | 1.16 <i>s</i> | 1.16 <i>s</i> | 1.16 <i>s</i> | 1.20 <i>s</i> | 1.21 <i>s</i> |
| 13 | 1.07 <i>s</i> | 1.06 <i>s</i> | 1.07 <i>s</i> | 0.96 <i>s</i> | 0.97 <i>s</i> |
| 14 | 1.48 <i>d</i> (1) | 1.47 <i>d</i> (0.9) | 1.48 <i>d</i> (1.1) | 1.55 <i>br s</i> | 1.57 <i>br s</i> |
| 15 | 4.96 <i>d</i> (2.1) | 4.96 <i>d</i> (2.1) | 4.96 <i>d</i> (2.1) | 9.42 <i>d</i> (1) | 9.44 <i>d</i> (0.9) |
| 15' | 4.90 <i>d</i> (2.1) | 4.90 <i>d</i> (2.1) | 4.90 <i>d</i> (2.1) | | |
| 3" | 8.03 <i>dd</i> (2; 8.9) | 7.99 <i>d</i> (8.8) | 7.66 <i>dd</i> (1.8; 8.3) | 7.98 <i>dd</i> (1.4; 8.7) | 7.60 <i>dd</i> (1.9; 8.4) |
| 4" | 7.46 <i>dt</i> (2; 8.8) | 6.91 <i>d</i> (8.8) | 6.96 <i>d</i> (8.3) | 7.41 <i>dt</i> (1.3; 8.6) | 6.93 <i>d</i> (8.4) |
| 5" | 7.60 <i>dt</i> (2; 8.8) | | | 7.53 <i>dt</i> (1.3; 8.6) | |
| 6" | 7.46 <i>dt</i> (2; 8.8) | 6.91 <i>d</i> (8.8) | | 7.41 <i>dt</i> (1.3; 8.6) | |
| 7" | 8.03 <i>dd</i> (2; 8.9) | 7.99 <i>d</i> (8.8) | 7.62 <i>d</i> (1.8) | 7.98 <i>dd</i> (1.4; 8.7) | 7.50 <i>d</i> (1.9) |
| OCH ₃ | | | 3.95 <i>s</i> | | 3.93 <i>s</i> |

those of **4** and **5**. Indeed, the ^1H NMR of **7** was only slightly different to the one recorded for **4**; these differences included the absence of an aldehydic proton signal, a signal for a more deshielded H-5 and a slightly shielded H-6 signal. The IR, ^{13}C NMR and mass spectra of **7** confirmed that the C-4 aldehyde group of **4** had been oxidized to a carboxyl group in **7**. 2D NMR homonuclear (COSY-45) and ^1H – ^{13}C heteronuclear (HETCOR) shift correlation experiments performed with **7** confirmed the ^1H and ^{13}C NMR assignments. Similarity of the chemical shift of H-5 in the ^1H NMR of **7** with that exhibited by **13**, a compound which was prepared from its $\Delta^4 Z$ isomer by photochemical transformation [15], suggested the same $\Delta^4 E$ configuration for **7**. Not only the $\Delta^4 E$ configuration but also identical stereochemistry for **4** and **7** was confirmed by correlation of their ^1H and ^{13}C NMR data. These assignments were supported by the oxidation of **4** to **7**. The acyl moieties of **6** and **7** were identified as angelate and benzoate, respectively.

The spectral data of the angelate (**8**) ($\text{C}_{20}\text{H}_{30}\text{O}_5$) and benzoate (**9**) ($\text{C}_{22}\text{H}_{28}\text{O}_5$) of $1\beta,10\alpha$ -epoxykurubaschic acid as well as the data for $1\alpha,10\beta$ -epoxykurubaschic acid benzoate (**10**) ($\text{C}_{22}\text{H}_{28}\text{O}_5$) indicated that all three were $1\alpha,10$ -epoxy analogues of the kurubaschic acid esters. While spectral similarity suggested the same stereochemistry for **8** and **9**, the spectral comparison indicated different stereochemistry for **10**. An angelate acyl group for **8** and benzoate for both **9** and **10** were assigned on the basis of their IR, MS, ^1H and ^{13}C NMR data. To elucidate the stereochemistry of these compounds, all ^1H NMR signals of **9** and **10** (diastereomeric pair) were identified by means of 2D NMR homonuclear COSY-45 experiments; then using NOESY experiments (Table 3) their stereochemistries were established as $1\alpha,10\beta$ -epoxy- and $1\beta,10\alpha$ -epoxykurubaschic acid benzoates, respectively. C-4 methyl analogues of these compounds with different acyl groups (**11** and **12**) were reported recently [16, 17], including X-ray crystallographic analysis. The ^1H and

(CDCl₃, TMS as int. standard, *J* in Hz in parentheses)

| | 6 (200 MHz) | 7 (500 MHz) | 8 (200 MHz) | 9 (500 MHz) | 10 (500 MHz) |
|------------------|-------------------------------|------------------------------------|-------------------------------------|-------------------------------------|----------------------------------|
| H-1 | 5.25 br <i>t</i> (7.6) | 5.27 br <i>t</i> (7.4) | 2.98 <i>dd</i> (3.4; 11.3) | 2.98 <i>dd</i> (3.4; 11.4) | 2.89 <i>dd</i> (4; 10.9) |
| H-2 | 2.30 <i>m</i> | 2.28 <i>m</i> | 2.25 <i>ddd</i> (1.5; 6.5; 14.4) | 2.24 <i>ddd</i> (3.1; 6.4; 14.1) | 2.45 <i>tt</i> (3.7; 13.7) |
| H-2' | | | 1.58 br <i>q</i> (14.4) | 1.57 <i>ddt</i> (1.5; 13; 14.4) | 1.47 <i>m</i> |
| H-3 | | 2.68 <i>ddd</i> (5.3; 10; 12.6) | 2.96 <i>dt</i> (1.4; 14.2) | 2.97 <i>dt</i> (1.4; 14.2) | 2.77 <i>dt</i> (3.3; 14.1) |
| H-3' | | 2.61 <i>td</i> (4.1; 12.6) | 2.70 br <i>dd</i> (6.3; 14.2) | 2.71 br <i>dd</i> (6.4; 14.2) | 2.69 <i>td</i> (3.7; 14.1) |
| H-5 | 6.93 <i>d</i> (10.6) | 6.98 <i>d</i> (10.6) | 6.88 <i>d</i> (10.6) | 6.94 <i>d</i> (10.9) | 6.88 <i>d</i> (10.5) |
| H-6 | 5.45 <i>d</i> (10.6) | 5.58 <i>d</i> (10.6) | 5.53 <i>d</i> (10.6) | 5.66 <i>d</i> (10.9) | 5.73 <i>d</i> (10.5) |
| H-7 | 1.31 <i>m</i> | | | 1.41 <i>m</i> | 1.47 <i>m</i> |
| | 1.25 <i>m</i> | | 1.38 <i>m</i> | | |
| H-7' | | 1.18 <i>m</i> | | | 1.22 <i>m</i> |
| H-8 | 1.63 <i>m</i> | | | 1.41 <i>m</i> | 1.68 <i>m</i> |
| | 1.48 <i>m</i> | | 1.38 <i>m</i> | | |
| H-8' | | 1.32 <i>m</i> | | 1.34 <i>m</i> | 1.28 <i>m</i> |
| H-9 | 2.21 br <i>d</i> (12.6) | 2.19 br <i>d</i> (12.6) | 1.82 br <i>dd</i> (11.6; 14.3) | 1.81 br <i>dd</i> (11.7; 12.4) | 2.07 <i>dd</i> (8.1; 12.7) |
| H-9' | 1.48 <i>dt</i> (2.1; 12.6) | 1.47 <i>dt</i> (2.1; 12.6) | 1.25 <i>m</i> | 1.22 <i>dd</i> (8.1; 12.4) | 0.83 <i>b dd</i> (11.6; 12.7) |
| H-12 | 1.07 <i>s</i> | 1.17 <i>s</i> | 1.10 <i>s</i> | 1.19 <i>s</i> | 1.16 <i>s</i> |
| H-13 | 0.89 <i>s</i> | 0.93 <i>s</i> | 0.95 <i>s</i> | 0.98 <i>s</i> | 0.98 <i>s</i> |
| H-14 | 1.61 br <i>s</i> | 1.61 <i>s</i> | 1.18 <i>s</i> | 1.17 <i>s</i> | 1.39 <i>s</i> |
| H-15 | | | | | |
| H-15' | | | | | |
| H-3" | 6.06 <i>qq</i> (1.4; 7.2) | 7.98 <i>dd</i> (1.4; 8.7) | 6.10 <i>qq</i> (1.4; 7.2) | 7.99 <i>dd</i> (1.4; 8.1) | 7.99 <i>dd</i> (1.3; 8.1) |
| H-4" | 1.96 <i>dq</i> (1.4; 7.2) | 7.41 <i>dt</i> (1.3; 8.6) | 1.98 <i>dq</i> (1.4; 7.2) | 7.42 <i>dt</i> (1.3; 8.1) | 7.43 <i>dt</i> (1.3; 8.2) |
| H-5" | 1.86 <i>t</i> (1.4) | 7.53 <i>dt</i> (1.3; 8.6) | 1.88 <i>t</i> (1.4) | 7.54 <i>dt</i> (1.3; 8.1) | 7.54 <i>dt</i> (1.3; 8.2) |
| H-6" | | 7.41 <i>dt</i> (1.3; 8.6) | | 7.42 <i>dt</i> (1.3; 8.1) | 7.43 <i>dt</i> (1.3; 8.2) |
| H-7" | | 7.98 <i>dd</i> (1.4; 8.7) | | 7.99 <i>dt</i> (1.4; 8.1) | 7.99 <i>dd</i> (1.3; 8.1) |
| OCH ₃ | | | | | |

¹³C NMR data correlation of **9** with that of **11** and **10** with that of **12**, especially for their similar structural fragments, clearly supported these assignments.

Humulenoid sesquiterpenes are relatively rare compounds in nature and the *cis* Δ^4 double bond conformational feature in the compounds described here distinguishes these compounds from the common type of all-*trans*-humulenes. Apparently, these *cis* Δ^4 double bond-containing compounds (α -apienes) are derived from *cis,trans*-farnesyl pyrophosphate [18] in contrast to the *trans,trans*-farnesyl pyrophosphate precursor of the latter type. Due to their *cis,trans*-farnesyl pyrophosphate precursor the *s-trans* 4(15), 5-diene conformation of servanol esters (1-3) (γ -apienes) is formed instead of *s-cis* 4(15), 5-diene conformation of common all-*trans*-humulene analogue γ -humulenes. It is of interest that to date all humulenoids containing *cis* Δ^4 double bonds or *s-trans* 4(15), 5-dienes were isolated from the Apiaceae; therefore,

we named this subclass as apienes (similar to the germacrane-heliangene relationship).

EXPERIMENTAL

Plant material. The plant material was collected from the Kurubaş Pass area between Van-Gürpınar (Eastern Anatolia) in June 1983. A voucher specimen is deposited in the Herbarium of Dicle University (DUF) (Herb. no. SAYA 83-167).

Isolation of compounds. Air-dried and finely cut root pieces of *F. haussknechtii* (13 g) were extracted with CH₂Cl₂ (100 ml) at room temp. for 20 min. Concentration of the CH₂Cl₂ extract *in vacuo* provided 1.1 g of a crude viscous oil. This oil was dissolved in Me₂CO (50 ml) and left overnight in a refrigerator. Following the removal of precipitated hydrocarbon mixtures by filtration, the solvent was removed *in vacuo* to yield 980 mg of a light yellow viscous oil. This material was chromatographed on a Sephadex LH-20 column (3 × 50 cm) packed in cyclohexane

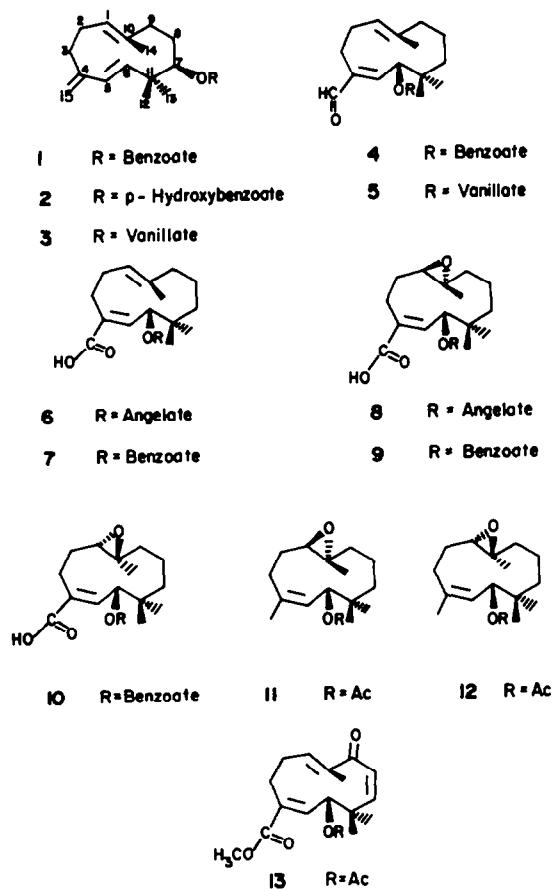


Table 3. NOEs with compounds 3, 4, 9 and 10

| Irrad. of | 3 NOE |
|------------------|---|
| Compound 3 | |
| H-5 | H-12, H-15 |
| H-6 | H-7, H-1, H-3 α |
| H-7 | H-6, H-9 α |
| H-12 | H-5 |
| H-13 | H-6, H-7 |
| H-14 | H-2 β |
| Compound 4 | |
| H-1 | H-6, H-7 α , H-3 α , H-9 α |
| H-5 | H-12, H-15 |
| H-6 | H-1, H-3 α , H-7 α , H-13 |
| H-14 | H-2 β |
| H-15 | H-5 |
| Compound 9 | |
| H-5 | H-12 |
| H-6 | H-1, H-3 α , H-13 |
| H-1 & 3 α | H-6, H-2 α |
| H-12 & 14 | H-5 |
| Compound 10 | |
| H-1 | H-7 β , H-9 β |
| H-5 | H-12 |
| H-6 | H-3 α , H-8 α |
| H-12 | H-5 |
| H-13 | H-6, H-8 α |
| H-14 | H-3 α , H-8 α , H-9 α |

Table 2. ^{13}C NMR spectral data for compounds 3, 4, 7 and 10 (CDCl₃, TMS)

| C | 3 (125.8 MHz) | 4 (22.6 MHz) | 7 (125.8 MHz) | 10 (125.8 MHz) |
|------------------|------------------|-----------------|------------------|-------------------|
| 1 | 126.9 <i>d</i> | 124.4 <i>d</i> | 123.9 <i>d</i> | 60.8 <i>d</i> |
| 2 | 30.2 <i>t</i> | 25.3 <i>t</i> | 26.0 <i>t</i> | 26.2 <i>t</i> |
| 3 | 31.5 <i>t</i> | 25.6 <i>t</i> | 27.7 <i>t</i> | 25.1 <i>t</i> |
| 4 | 148.3 <i>s</i> | 143.0 <i>s</i> | 136.1 <i>s</i> | 136.0 <i>s</i> |
| 5 | 138.5 <i>d</i> | 143.0 <i>d</i> | 138.3 <i>d</i> | 138.2 <i>d</i> |
| 6 | 128.5 <i>d</i> | 72.8 <i>d</i> | 73.2 <i>d</i> | 76.7 <i>d</i> |
| 7 | 83.6 <i>d</i> | 35.8 <i>t</i> | 35.6 <i>t</i> | 37.3 <i>t</i> |
| 8 | 30.2 <i>t</i> | 23.8 <i>t</i> | 23.7 <i>t</i> | 19.8 <i>t</i> |
| 9 | 40.9 <i>t</i> | 35.8 <i>t</i> | 35.9 <i>t</i> | 38.3 <i>t</i> |
| 10 | 135.5 <i>s</i> | 136.8 <i>s</i> | 136.9 <i>s</i> | 60.6 <i>s</i> |
| 11 | 37.7 <i>s</i> | 37.8 <i>s</i> | 37.8 <i>s</i> | 37.7 <i>s</i> |
| 12 | 26.0 <i>q</i> | 24.3 <i>q</i> | 24.3 <i>q</i> | 24.5 <i>q</i> |
| 13 | 26.0 <i>q</i> | 22.9 <i>q</i> | 22.8 <i>q</i> | 22.9 <i>q</i> |
| 14 | 17.0 <i>q</i> | 19.4 <i>q</i> | 19.2 <i>q</i> | 16.6 <i>q</i> |
| 15 | 114.7 <i>t</i> | 218.1 <i>d</i> | 173.4 <i>s</i> | 171.7 <i>s</i> |
| 1' | 166.0 <i>s</i> | 165.5 <i>s</i> | 165.5 <i>s</i> | 165.8 <i>s</i> |
| 2' | 122.9 <i>s</i> | 130.2 <i>s</i> | 130.2 <i>s</i> | 130.0 <i>s</i> |
| 3' | 124.0 <i>d</i> | 129.7 <i>d</i> | 129.6 <i>d</i> | 129.6 <i>d</i> |
| 4' | 111.8 <i>d</i> | 128.5 <i>d</i> | 128.3 <i>d</i> | 128.5 <i>d</i> |
| 5' | 148.3 <i>s</i> | 133.2 <i>d</i> | 132.9 <i>d</i> | 133.2 <i>d</i> |
| 6' | 150.0 <i>s</i> | 128.5 <i>d</i> | 128.3 <i>d</i> | 128.5 <i>d</i> |
| 7' | 114.1 <i>d</i> | 129.7 <i>d</i> | 129.6 <i>d</i> | 129.6 <i>d</i> |
| OCH ₃ | 56.1 <i>q</i> | | | |

$-\text{CH}_2\text{Cl}_2$ –EtOH (7:4:1) and prep. TLC (1.5–2 mm thickness, silica gel developed with cyclohexane–EtOAc mixtures, 4:1, 7:3 or 3:2) was used for final purification of compounds.

Fervanol benzoate (1). Gum (21 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm^{-1} : 3070, 2970, 2875, 1720, 1602, 1585, 1450, 1270, 1215, 1040, 880, 830, 710, 688, 675. EIMS (probe, 70 eV) m/z (rel. int.): 324 [M]⁺ (1.2), 202 [M–benzoic acid]⁺ (12.1), 187 (5.6), 159 (9.4), 122 [benzoic acid]⁺ (60), 105 [benzoate]⁺ (100).

Fervanol p-hydroxybenzoate (2). Gum (34 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm^{-1} : 3360, 2970, 2940, 2870, 1680, 1610, 1595, 1512, 1450, 1365, 1280, 1220, 1160, 1100, 880, 850, 830, 770, 700. EIMS (probe, 70 eV) m/z (rel. int.): 340 [M]⁺ (0.8), 202 [M–p-hydroxybenzoic acid]⁺ (9.4), 187 (4.9), 159 (8.8), 138 [p-hydroxybenzoic acid]⁺ (40), 121 [p-hydroxybenzoate]⁺ (100).

Fervanol vanillate (3). Gum (9 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm^{-1} : 3400, 3080, 2975, 2938, 2870, 1710, 1610(sh), 1600, 1515, 1460, 1450, 1428, 1283, 1220, 1105, 1030, 880, 830, 785, 762, 725. EIMS (probe, 70 eV) m/z (rel. int.): 370 [M]⁺ (4), 202 [M–vanillic acid]⁺ (13.9), 187 (11.6), 168 [vanillic acid]⁺ (18), 159 (15.8), 151 [vanillate]⁺ (100).

Kurubasch aldehyde benzoate (4). Gum (45 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm^{-1} : 3060, 2970, 2930, 2865, 2710, 1720, 1695, 1602, 1586, 1490, 1450, 1318, 1270, 1110, 1098, 1028, 950, 935, 800, 710, 688, 672. EIMS (probe, 70 eV) m/z (rel. int.): 340 [M]⁺ (6.9), 218 [M–benzoic acid]⁺ (14.8), 203 (13.6), 189 (8.6), 175 (10), 122 [benzoic acid]⁺ (16.6), 105 [benzoate]⁺ (100).

Oxidation of 4. Kurubasch aldehyde benzoate (4) (10 mg) was dissolved in Et₂O (3 ml), chromic acid soln (0.3 ml) [20] was added dropwise and the soln stirred at room temp. for 1 hr. The Et₂O layer was separated and washed with NaHCO₃ soln (1%), dried over anhyd. Na₂SO₄ and evapd *in vacuo* to yield 7 mg gum. Spectral properties of the product were found to be identical with 7.

Kurubasch aldehyde vanillate (5). Gum (11 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm^{-1} : 3400, 3080, 2970, 2940, 2870, 2715, 1710(sh), 1692, 1610(sh), 1600, 1512, 1460, 1430, 1388, 1370, 1280, 1215, 1100, 1032, 958, 878, 785, 762, 725. EIMS (probe, 70 eV) m/z (rel. int.): 386 [M]⁺ (3.3), 218 [M–vanillic acid]⁺ (5.9), 203 (2.3), 189 (3.1), 175 (4), 168 [vanillic acid]⁺ (15.9), 151 [vanillate]⁺ (100).

Kurubaschic acid angelate (6). Gum (25 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm^{-1} : 3100(sh), 2970, 2940, 2880, 2670, 2520(sh), 1715(sh), 1695, 1640, 1510, 1458, 1385, 1368, 1355, 1270, 1230, 1150, 1040, 960, 938, 845, 795. EIMS (probe, 70 eV) m/z (rel. int.): 334 [M]⁺ (1.1), 234 [M–angelic acid]⁺ (13), 219 (15.9), 206 (10.1), 191 (13.2), 149 (67.8), 105 (97), 100 [angelic acid]⁺ (68.6), 83 [angelate]⁺ (100).

Kurubaschic acid benzoate (7). Gum (230 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm^{-1} : 3100(sh), 3065, 2970, 2940, 2875, 2660, 2520(sh), 1720, 1692, 1640, 1602, 1585, 1512, 1450, 1388, 1370, 1320, 1270, 1210, 1108, 1070, 1026, 1002, 952, 938, 880, 796, 710, 688, 675. EIMS (probe, 70 eV) m/z (rel. int.): 356 [M]⁺ (0.14), 234 [M–benzoic acid]⁺ (32), 219 (33.9), 206 (5.7), 191 (24.7), 149 (38.5), 122 [benzoic acid]⁺ (82.4), 105 [benzoate]⁺ (100). CIMS (methane, probe, 70 eV) m/z (rel. int.): 355 [M–1]⁺ (5.9), 339 [M+1–H₂O]⁺ (6.4), 251 [M–1–benzoate+H]⁺ (9.8), 235 [M+1–benzoic acid]⁺ (100), 233 [M–1–benzoic acid]⁺ (62.5), 217 (43.2), 205 (20.3), 189 (48), 123 [benzoic acid+H]⁺ (59.8), 105 [benzoate]⁺ (51.5).

$\beta,10\alpha$ -Epoxykurubaschic acid angelate (8). Gum (12 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm^{-1} : 3200(sh), 2975, 2930, 2870, 2660, 2520(sh), 1715, 1695, 1645, 1510, 1453, 1385, 1270, 1260, 1230, 1150, 1040, 962, 862, 780, 710. EIMS (probe, 70 eV) m/z (rel. int.): 350 [M]⁺ (0.1), 250 [M–angelic acid]⁺ (4.4), 235 (2.7), 232 (4.2), 205 (5.2), 191 (6), 189 (6.6), 165 (12.9), 151 (17.7), 149 (17.6), 121 (19.3), 105 (28.9), 100 [angelic acid]⁺ (18.4), 83 [angelate]⁺ (100).

$\beta,10\alpha$ -Epoxykurubaschic acid benzoate (9). Gum (14 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm^{-1} : 3200(sh), 3060, 2970, 2940, 2875, 2660, 2520(sh),

1720, 1693, 1642, 1602, 1585, 1490, 1450, 1385, 1370, 1320, 1270, 1110, 1070, 1025, 955, 770, 710, 685, 677. EIMS (probe, 70 eV) m/z (rel. int.): 372 [M]⁺ (0.12), 250 [M–benzoic acid]⁺ (2.5), 235 (2.1), 232 (3.9), 205 (2), 191 (3.7), 189 (4.3), 165 (6.6), 151 (5.1), 122 [benzoic acid]⁺ (82.2), 105 [benzoate]⁺ (100). CIMS (methane, probe, 70 eV) m/z (rel. int.): 373 [M+1]⁺ (1.1), 371 [M–1]⁺ (1.3), 251 [M+1–benzoic acid]⁺ (20.6), 233 [M+1–benzoic acid–H₂O]⁺ (100), 223 (6.7), 215 (16.2), 205 (39.3), 193 (8.7), 187 (19.8), 177 (11.4), 123 [benzoic acid+H]⁺ (18.2), 105 [benzoate]⁺ (25.2).

$\alpha,10\beta$ -Epoxykurubaschic acid benzoate (10). Gum (8.5 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm^{-1} : 3200(sh), 3070, 2970, 2940, 2870, 2660, 2530(sh), 1720, 1695, 1645, 1602, 1588, 1490(sh), 1465, 1450, 1388, 1370, 1270, 1178, 1110, 1070, 1050, 1028, 960, 865, 775, 710, 688, 678. EIMS (probe, 70 eV) m/z (rel. int.): 272 [M]⁺ (0.3), 250 [M–benzoic acid]⁺ (1.8), 235 (1.4), 191 (3.1), 189 (3.9), 165 (6.4), 151 (3.7) 122 [benzoic acid]⁺ (56), 105 [benzoate]⁺ (100).

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REFERENCES

1. Miski, M., Ulubelen, A. and Mabry, T. J. (1983) *Phytochemistry* **22**, 2231.
2. Miski, M., Ulubelen, A., Mabry, T. J., Watson, W. H., Vickovic, I. and Holub, M. (1984) *Tetrahedron* **40**, 5197.
3. Miski, M. and Mabry, T. J. (1985) *Phytochemistry* **24**, 1735.
4. Miski, M. and Mabry, T. J. (1986) *Phytochemistry* **25**, 1673.
5. Saidkhodzhaev, A. I. (1979) *Khim. Prir. Soedin.* 137.
6. Sagitdinova, G. V. and Saidkhodzhaev, A. I. (1977) *Khim. Prir. Soedin.* 790.
7. Sagitdinova, G. V., Saidkhodzhaev, A. I. and Malikov, V. M. (1978) *Khim. Prir. Soedin.* 809.
8. Sagitdinova, G. V., Saidkhodzhaev, A. I. and Malikov, V. M. (1979) *Khim. Prir. Soedin.* 864.
9. Buzhanova, K., Saidkhodzhaev, A. I. and Malikov, V. M. (1978) *Khim. Prir. Soedin.* 407.
10. Buzhanova, K., Saidkhodzhaev, A. I. and Malikov, V. M. (1978) *Khim. Prir. Soedin.* 576.
11. Sagitdinova, G. V., Saidkhodzhaev, A. I. and Malikov, V. M. (1982) *Khim. Prir. Soedin.* 721.
12. Golovina, L. A. and Saidkhodzhaev, A. I. (1977) *Khim. Prir. Soedin.* 796.
13. Bohlmann, F., Zdero, C. and Grenz, M. (1974) *Chem. Ber.* **107**, 3928.
14. Bohlmann, F., Zdero, C., King, R. M. and Robinson, H. (1982) *Phytochemistry* **21**, 147.
15. El Dahmy, S., Jakupovic, J., Bohlmann, F. and Sarg, T. M. (1985) *Tetrahedron* **41**, 309.
16. Itokawa, H., Matsumoto, H., Mihashi, S. and Iitika, Y. (1983) *Chem. Letters* 1581.
17. Itokawa, H., Matsumoto, H., Mihashi, S., Iitika, Y., Kasuya, A. and Itai, A. (1985) *Chem. Pharm. Bull.* **33**, 2204.
18. Hendrickson, J. B. (1959) *Tetrahedron* **7**, 82.
19. Sasaki, S., Itagaki, Y., Moriyama, H., Nakanishi, K., Watanabe, E. and Aoyama, T. (1966) *Tetrahedron Letters* 1623.
20. Brown, H. C., Garg, C. P. and Liu, K.-T. (1971) *J. Org. Chem.* **36**, 387.