

APIENE ESTERS FROM *FERULA HAUSSKNECHTII*

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Key Word Index—*Ferula haussknechtii*; Apiaceae; sesquiterpene esters; apienes.

Abstract—Ten new apiene-type humulenoid esters were isolated from the root of *Ferula haussknechtii*. The structures were elucidated by spectroscopic methods.

INTRODUCTION

In continuation of our investigation of *Ferula* species which are indigenous to the Anatolia region of Turkey [1–4], we investigated *F. haussknechtii* Wolff. ex Rech. *Ferula* species are known to be a rich source of different classes of sesquiterpenes such as daucanes, germacranes, eudasmanes, guaianes, elemanes, humulenes and himacalenes [5]. Although the occurrence of humulenes is relatively rare in the genus *Ferula*, α -humulenes from *F. juniperina* [6–8], *F. xeromorpha* [9, 10] and *F. tschatalensis* [11], as well as γ -humulenes from *F. ceratophylla* [12], have been reported previously.

RESULTS AND DISCUSSION

The dichloromethane extract of the root of *F. haussknechtii* afforded ten new sesquiterpene esters in small amounts; their structures were elucidated by spectral techniques, especially stereochemical data obtained with the relatively new techniques such as 2D NMR homo- and heteronuclear COSY and high resolution NOESY experiments.

The IR spectra of the benzoate (1) ($C_{22}H_{28}O_2$), *p*-hydroxybenzoate (2) ($C_{22}H_{28}O_3$) and vanillate (3) ($C_{23}H_{30}O_4$) derivatives of fervalol all exhibited absorptions for an aromatic acyl group. The 1H NMR (Table 1), ^{13}C NMR (Table 2) and MS data for 1, 2 and 3 confirmed the presence of benzoate, *p*-hydroxybenzoate and vanillate acyl groups, respectively, as well as the same sesquiterpene nucleus for all three. A total of four degrees of unsaturation were assigned for this nucleus based on the ^{13}C NMR and mass spectral data. The 1H NMR and ^{13}C NMR spectra of 1–3 indicated the presence of an exocyclic methylene group (δ 4.96, 4.90, each 1H, *d* and 148.3, *s*; 114.7, *t*), (a trisubstituted double bond (δ 5.32, 1H, *t*; 1.48, 3H, *d* and 126.9, *d*; 135.5, *s*) and a *trans* disubstituted double bond (δ 6.04 and 5.59, each 1H, *d*, *J* = 16.1 Hz; 138.5, *d* and 128.3, *d*), as well as two geminal methyl groups (δ 1.16 and 1.07, each 3H, *s*; both at 26.0, *q*). The presence of these three double bonds indicated a monocyclic structure for all three compounds; moreover, the presence of two geminal methyl groups instead of an isopropyl group clearly demonstrated the γ -humulenoid-type structure for 1–3. The location of the aromatic acyl

groups and the structural relationship of these three compounds was deduced from 2D NMR homonuclear COSY-45 experiments. Nevertheless, the spectral data did not establish all stereochemical matters. Interestingly, in the 1H NMR spectra of 1–3 the exocyclic methylene proton signals appear as two doublets, whereas the same signals in the spectra of all previously known γ -humulenes except for the compound obtained from *F. ceratophylla*, which probably has the same stereochemical features as 1–3, were reported as broad singlets [13, 14]. The 2D NMR homonuclear COSY-45 experiment confirmed the presence of geminal coupling between the exocyclic methylene protons but no other long range couplings with these protons. This feature is probably indicative of a different conformation of 1–3 relative to previously reported γ -humulenes. Further information regarding the stereochemistry of 1–3 was obtained by a series of NOESY experiments. The 1H NMR NOESY experiments of 3 verified an *s-trans* for the conjugated 4(15),5-diene system and β orientation for the C-7 acyl group.

The IR spectra of the benzoate (4) ($C_{22}H_{28}O_3$) and vanillate (5) ($C_{23}H_{30}O_4$) of kurubasch aldehyde showed an α,β -unsaturated aldehyde (1695 cm^{-1}) and aromatic acyl group absorptions. The aromatic acyl groups of 4 and 5 were assigned as benzoate and vanillate, respectively, on the basis of their 1H , ^{13}C NMR and mass spectral data. Similarity of their spectra, except for the acyl group signals, confirmed the same sesquiterpene skeleton in both 4 and 5. In addition to the other spectral data, an α -humulenoid structure for 4 and 5 was deduced from the 2D NMR homonuclear COSY-45 experiments. In the 1H NMR spectra of 4 and 5 the downfield position of the H-5 signal (δ 6.50, 1H, *d*) and the presence of an aldehyde proton signal (δ 9.4, 1H, *d*) instead of a second vinylic methyl signal expected for an α -humulene, clearly indicated that in 4 and 5 the C-4 methyl of α -humulenes had been replaced with an aldehyde group. Furthermore, the doublet at δ 5.7 (1H), which coupled with the former H-5 signal, at δ 6.50, confirmed the location of the aromatic acyl groups to be C-6 in both compounds. The NOESY experiments of 4 (Table 3) showed that both the $\Delta^{1,10}$ and Δ^4 double bonds had the *E* configuration and that the stereochemistry of the acyl group at C-6 is β .

Kurubaschic acid angelate (6) ($C_{20}H_{30}O_4$) and benzoate (7) ($C_{22}H_{28}O_4$) exhibited similar spectral data to

Table 1. ^1H NMR spectra data of compounds 1–10

H	1 (200 MHz)	2 (200 MHz)	3 (500 MHz)	4 (500 MHz)	5 (200 MHz)
1	5.32 <i>br t</i> (8.5)	5.31 <i>br t</i> (8.4)	5.32 <i>br t</i> (8.5)	5.32 <i>br dd</i> (6.3; 5.8)	5.24 <i>br t</i> (7.6)
2	2.22 <i>m</i>	2.21 <i>m</i>	2.22 <i>m</i>	2.28 <i>m</i>	2.26 <i>m</i>
2'	2.33 <i>m</i>	2.32 <i>m</i>	2.33 <i>m</i>	2.19 <i>m</i>	
3	2.38 <i>m</i>	2.36 <i>m</i>	2.38 <i>m</i>	2.61 <i>td</i> (4.1; 12.6)	2.60 <i>m</i>
3'				2.54 <i>dt</i> (3.8; 12.6)	
5	6.04 <i>d</i> (16.1)	6.04 <i>d</i> (16.1)	6.04 <i>d</i> (16.1)	6.49 <i>d</i> (10.3)	6.50 <i>d</i> (10.3)
6	5.59 <i>d</i> (16.1)	5.58 <i>d</i> (16.1)	5.59 <i>d</i> (16.1)	5.72 <i>d</i> (10.3)	5.70 <i>d</i> (10.3)
7				1.33 <i>m</i>	
7'	4.68 <i>br d</i> (6.9)	4.68 <i>br d</i> (6.8)	4.68 <i>br d</i> (6.9)	1.20 <i>m</i>	1.38 <i>m</i>
8	2.20 <i>m</i>	2.19 <i>m</i>	2.24 <i>m</i>	1.64 <i>m</i>	1.60 <i>m</i>
8'	1.91 <i>m</i>	1.90 <i>m</i>	1.92 <i>m</i>	1.36 <i>m</i>	1.38 <i>m</i>
9	2.17 <i>m</i>	2.17 <i>m</i>	2.17 <i>m</i>	2.20 <i>m</i>	2.21 <i>m</i>
9'	1.58 <i>m</i>	1.58 <i>m</i>	1.58 <i>m</i>	1.46 <i>dt</i> (2.1; 13)	1.48 <i>dt</i> (2.1; 13)
12	1.16 <i>s</i>	1.16 <i>s</i>	1.16 <i>s</i>	1.20 <i>s</i>	1.21 <i>s</i>
13	1.07 <i>s</i>	1.06 <i>s</i>	1.07 <i>s</i>	0.96 <i>s</i>	0.97 <i>s</i>
14	1.48 <i>d</i> (1)	1.47 <i>d</i> (0.9)	1.48 <i>d</i> (1.1)	1.55 <i>br s</i>	1.57 <i>br s</i>
15	4.96 <i>d</i> (2.1)	4.96 <i>d</i> (2.1)	4.96 <i>d</i> (2.1)		
15'	4.90 <i>d</i> (2.1)	4.90 <i>d</i> (2.1)	4.90 <i>d</i> (2.1)	9.42 <i>d</i> (1)	9.44 <i>d</i> (0.9)
3"	8.03 <i>dd</i> (2; 8.9)	7.99 <i>d</i> (8.8)	7.66 <i>dd</i> (1.8; 8.3)	7.98 <i>dd</i> (1.4; 8.7)	7.60 <i>dd</i> (1.9; 8.4)
4"	7.46 <i>dt</i> (2; 8.8)	6.91 <i>d</i> (8.8)	6.96 <i>d</i> (8.3)	7.41 <i>dt</i> (1.3; 8.6)	6.93 <i>d</i> (8.4)
5"	7.60 <i>dt</i> (2; 8.8)			7.53 <i>dt</i> (1.3; 8.6)	
6"	7.46 <i>dt</i> (2; 8.8)	6.91 <i>d</i> (8.8)		7.41 <i>dt</i> (1.3; 8.6)	
7"	8.03 <i>dd</i> (2; 8.9)	7.99 <i>d</i> (8.8)	7.62 <i>d</i> (1.8)	7.98 <i>dd</i> (1.4; 8.7)	7.50 <i>d</i> (1.9)
OCH ₃			3.95 <i>s</i>		3.93 <i>s</i>

those of 4 and 5. Indeed, the ^1H NMR of 7 was only slightly different to the one recorded for 4; these differences included the absence of an aldehydic proton signal, a signal for a more deshielded H-5 and a slightly shielded H-6 signal. The IR, ^{13}C NMR and mass spectra of 7 confirmed that the C-4 aldehyde group of 4 had been oxidized to a carboxyl group in 7. 2D NMR homonuclear (COSY-45) and ^1H – ^{13}C heteronuclear (HETCOR) shift correlation experiments performed with 7 confirmed the ^1H and ^{13}C NMR assignments. Similarity of the chemical shift of H-5 in the ^1H NMR of 7 with that exhibited by 13, a compound which was prepared from its $\Delta^4 Z$ isomer by photochemical transformation [15], suggested the same $\Delta^4 E$ configuration for 7. Not only the $\Delta^4 E$ configuration but also identical stereochemistry for 4 and 7 was confirmed by correlation of their ^1H and ^{13}C NMR data. These assignments were supported by the oxidation of 4 to 7. The acyl moieties of 6 and 7 were identified as angelate and benzoate, respectively.

The spectral data of the angelate (8) ($\text{C}_{20}\text{H}_{30}\text{O}_5$) and benzoate (9) ($\text{C}_{22}\text{H}_{28}\text{O}_5$) of $1\beta,10\alpha$ -epoxykurubaschic acid as well as the data for $1\alpha,10\beta$ -epoxykurubaschic acid benzoate (10) ($\text{C}_{22}\text{H}_{28}\text{O}_5$) indicated that all three were 1, 10-epoxy analogues of the kurubaschic acid esters. While spectral similarity suggested the same stereochemistry for 8 and 9, the spectral comparison indicated different stereochemistry for 10. An angelate acyl group for 8 and benzoate for both 9 and 10 were assigned on the basis of their IR, MS, ^1H and ^{13}C NMR data. To elucidate the stereochemistry of these compounds, all ^1H NMR signals of 9 and 10 (diastereomeric pair) were identified by means of 2D NMR homonuclear COSY-45 experiments; then using NOESY experiments (Table 3) their stereochemistries were established as $1\alpha, 10\beta$ -epoxy- and $1\beta, 10\alpha$ -epoxykurubaschic acid benzoates, respectively. C-4 methyl analogues of these compounds with different acyl groups (11 and 12) were reported recently [16, 17], including X-ray crystallographic analysis. The ^1H and

(CDCl₃, TMS as int. standard, *J* in Hz in parentheses)

	6 (200 MHz)	7 (500 MHz)	8 (200 MHz)	9 (500 MHz)	10 (500 MHz)
H-1	5.25 <i>br t</i> (7.6)	5.27 <i>br t</i> (7.4)	2.98 <i>dd</i> (3.4; 11.3)	2.98 <i>dd</i> (3.4; 11.4)	2.89 <i>dd</i> (4; 10.9)
H-2			2.25 <i>ddd</i>	2.24 <i>ddd</i>	2.45 <i>tt</i>
	2.30 <i>m</i>	2.28 <i>m</i>	(1.5; 6.5; 14.4)	(3.1; 6.4; 14.1)	(3.7; 13.7)
H-2'			1.58 <i>br q</i> (14.4)	1.57 <i>ddt</i> (1.5; 13; 14.4)	1.47 <i>m</i>
H-3		2.68 <i>ddd</i> (5.3; 10; 12.6)	2.96 <i>dt</i> (1.4; 14.2)	2.97 <i>dt</i> (1.4; 14.2)	2.77 <i>dt</i> (3.3; 14.1)
H-3'	2.63 <i>m</i>	2.61 <i>td</i> (4.1; 12.6)	2.70 <i>br dd</i> (6.3; 14.2)	2.71 <i>br dd</i> (6.4; 14.2)	2.69 <i>td</i> (3.7; 14.1)
H-5	6.93 <i>d</i> (10.6)	6.98 <i>d</i> (10.6)	6.88 <i>d</i> (10.6)	6.94 <i>d</i> (10.9)	6.88 <i>d</i> (10.5)
H-6	5.45 <i>d</i> (10.6)	5.58 <i>d</i> (10.6)	5.53 <i>d</i> (10.6)	5.66 <i>d</i> (10.9)	5.73 <i>d</i> (10.5)
H-7		1.31 <i>m</i>			1.47 <i>m</i>
	1.25 <i>m</i>		1.38 <i>m</i>	1.41 <i>m</i>	
H-7'		1.18 <i>m</i>			1.22 <i>m</i>
H-8		1.63 <i>m</i>		1.41 <i>m</i>	1.68 <i>m</i>
	1.48 <i>m</i>		1.38 <i>m</i>		
H-8'		1.32 <i>m</i>		1.34 <i>m</i>	1.28 <i>m</i>
H-9	2.21 <i>br d</i> (12.6)	2.19 <i>br d</i> (12.6)	1.82 <i>br dd</i> (11.6; 14.3)	1.81 <i>br dd</i> (11.7; 12.4)	2.07 <i>dd</i> (8.1; 12.7)
H-9'	1.48 <i>dt</i> (2.1; 12.6)	1.47 <i>dt</i> (2.1; 12.6)	1.25 <i>m</i>	1.22 <i>dd</i> (8.1; 12.4)	0.83 <i>b dd</i> (11.6; 12.7)
H-12	1.07 <i>s</i>	1.17 <i>s</i>	1.10 <i>s</i>	1.19 <i>s</i>	1.16 <i>s</i>
H-13	0.89 <i>s</i>	0.93 <i>s</i>	0.95 <i>s</i>	0.98 <i>s</i>	0.98 <i>s</i>
H-14	1.61 <i>br s</i>	1.61 <i>s</i>	1.18 <i>s</i>	1.17 <i>s</i>	1.39 <i>s</i>
H-15					
H-15'					
H-3"	6.06 <i>qq</i> (1.4; 7.2)	7.98 <i>dd</i> (1.4; 8.7)	6.10 <i>qq</i> (1.4; 7.2)	7.99 <i>dd</i> (1.4; 8.1)	7.99 <i>dd</i> (1.3; 8.1)
H-4"	1.96 <i>dq</i> (1.4; 7.2)	7.41 <i>dt</i> (1.3; 8.6)	1.98 <i>dq</i> (1.4; 7.2)	7.42 <i>dt</i> (1.3; 8.1)	7.43 <i>dt</i> (1.3; 8.2)
H-5"	1.86 <i>t</i> (1.4)	7.53 <i>dt</i> (1.3; 8.6)	1.88 <i>t</i> (1.4)	7.54 <i>dt</i> (1.3; 8.1)	7.54 <i>dt</i> (1.3; 8.2)
H-6"		7.41 <i>dt</i> (1.3; 8.6)		7.42 <i>dt</i> (1.3; 8.1)	7.43 <i>dt</i> (1.3; 8.2)
H-7"		7.98 <i>dd</i> (1.4; 8.7)		7.99 <i>dt</i> (1.4; 8.1)	7.99 <i>dd</i> (1.3; 8.1)
OCH ₃					

¹³CNMR data correlation of 9 with that of 11 and 10 with that of 12, especially for their similar structural fragments, clearly supported these assignments.

Humulenoid sesquiterpenes are relatively rare compounds in nature and the *cis* Δ⁴ double bond conformational feature in the compounds described here distinguishes these compounds from the common type of all-*trans*-humulenes. Apparently, these *cis* Δ⁴ double bond-containing compounds (α-apienes) are derived from *cis,trans*-farnesyl pyrophosphate [18] in contrast to the *trans,trans*-farnesyl pyrophosphate precursor of the latter type. Due to their *cis,trans*-farnesyl pyrophosphate precursor the *s-trans* 4(15), 5-diene conformation of farnanol esters (1–3) (γ-apienes) is formed instead of *s-cis* 4(15), 5-diene conformation of common all-*trans*-humulene analogue γ-humulenes. It is of interest that to date all humulenoids containing *cis* Δ⁴ double bonds or *s-trans* 4(15), 5-dienes were isolated from the Apiaceae; therefore,

we named this subclass as apienes (similar to the germacrane–heliangene relationship).

EXPERIMENTAL

Plant material. The plant material was collected from the Kurubaş Pass area between Van-Gürpınar (Eastern Anatolia) in June 1983. A voucher specimen is deposited in the Herbarium of Dicle University (DUF) (Herb. no. SAYA 83–167).

Isolation of compounds. Air-dried and finely cut root pieces of *F. haussknechtii* (13 g) were extracted with CH₂Cl₂ (100 ml) at room temp. for 20 min. Concentration of the CH₂Cl₂ extract *in vacuo* provided 1.1 g of a crude viscous oil. This oil was dissolved in Me₂CO (50 ml) and left overnight in a refrigerator. Following the removal of precipitated hydrocarbon mixtures by filtration, the solvent was removed *in vacuo* to yield 980 mg of a light yellow viscous oil. This material was chromatographed on a Sephadex LH-20 column (3 × 50 cm) packed in cyclohexane

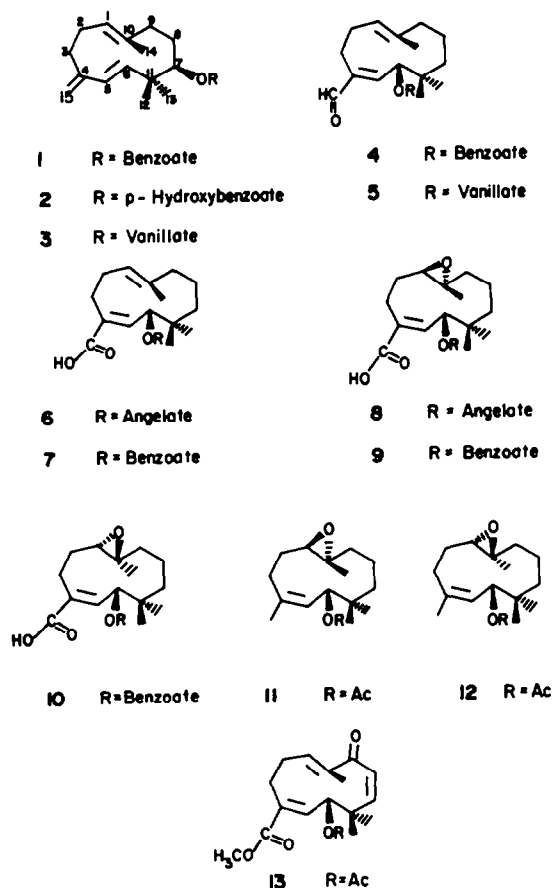


Table 3. NOEs with compounds 3, 4, 9 and 10

Irrad. of	3 NOE
Compound 3	
H-5	H-12, H-15
H-6	H-7, H-1, H-3 α
H-7	H-6, H-9 α
H-12	H-5
H-13	H-6, H-7
H-14	H-2 β
Compound 4	
H-1	H-6, H-7 α , H-3 α , H-9 α
H-5	H-12, H-15
H-6	H-1, H-3 α , H-7 α , H-13
H-14	H-2 β
H-15	H-5
Compound 9	
H-5	H-12
H-6	H-1, H-3 α , H-13
H-1 & 3 α	H-6, H-2 α
H-12 & 14	H-5
Compound 10	
H-1	H-7 β , H-9 β
H-5	H-12
H-6	H-3 α , H-8 α
H-12	H-5
H-13	H-6, H-8 α
H-14	H-3 α , H-8 α , H-9 α

Table 2. ^{13}C NMR spectral data for compounds 3, 4, 7 and 10 (CDCl_3 , TMS)

C	3 (125.8 MHz)	4 (22.6 MHz)	7 (125.8 MHz)	10 (125.8 MHz)
1	126.9 <i>d</i>	124.4 <i>d</i>	123.9 <i>d</i>	60.8 <i>d</i>
2	30.2 <i>t</i>	25.3 <i>t</i>	26.0 <i>t</i>	26.2 <i>t</i>
3	31.5 <i>t</i>	25.6 <i>t</i>	27.7 <i>t</i>	25.1 <i>t</i>
4	148.3 <i>s</i>	143.0 <i>s</i>	136.1 <i>s</i>	136.0 <i>s</i>
5	138.5 <i>d</i>	143.0 <i>d</i>	138.3 <i>d</i>	138.2 <i>d</i>
6	128.5 <i>d</i>	72.8 <i>d</i>	73.2 <i>d</i>	76.7 <i>d</i>
7	83.6 <i>d</i>	35.8 <i>t</i>	35.6 <i>t</i>	37.3 <i>t</i>
8	30.2 <i>t</i>	23.8 <i>t</i>	23.7 <i>t</i>	19.8 <i>t</i>
9	40.9 <i>t</i>	35.8 <i>t</i>	35.9 <i>t</i>	38.3 <i>t</i>
10	135.5 <i>s</i>	136.8 <i>s</i>	136.9 <i>s</i>	60.6 <i>s</i>
11	37.7 <i>s</i>	37.8 <i>s</i>	37.8 <i>s</i>	37.7 <i>s</i>
12	26.0 <i>q</i>	24.3 <i>q</i>	24.3 <i>q</i>	24.5 <i>q</i>
13	26.0 <i>q</i>	22.9 <i>q</i>	22.8 <i>q</i>	22.9 <i>q</i>
14	17.0 <i>q</i>	19.4 <i>q</i>	19.2 <i>q</i>	16.6 <i>q</i>
15	114.7 <i>t</i>	218.1 <i>d</i>	173.4 <i>s</i>	171.7 <i>s</i>
1'	166.0 <i>s</i>	165.5 <i>s</i>	165.5 <i>s</i>	165.8 <i>s</i>
2'	122.9 <i>s</i>	130.2 <i>s</i>	130.2 <i>s</i>	130.0 <i>s</i>
3'	124.0 <i>d</i>	129.7 <i>d</i>	129.6 <i>d</i>	129.6 <i>d</i>
4'	111.8 <i>d</i>	128.5 <i>d</i>	128.3 <i>d</i>	128.5 <i>d</i>
5'	148.3 <i>s</i>	133.2 <i>d</i>	132.9 <i>d</i>	133.2 <i>d</i>
6'	150.0 <i>s</i>	128.5 <i>d</i>	128.3 <i>d</i>	128.5 <i>d</i>
7'	114.1 <i>d</i>	129.7 <i>d</i>	129.6 <i>d</i>	129.6 <i>d</i>
OCH ₃	56.1 <i>q</i>			

—CH₂Cl₂—EtOH (7:4:1) and prep. TLC (1.5–2 mm thickness, silica gel developed with cyclohexane—EtOAc mixtures, 4:1, 7:3 or 3:2) was used for final purification of compounds.

Fervanol benzoate (1). Gum (21 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm⁻¹: 3070, 2970, 2940, 2875, 1720, 1602, 1585, 1450, 1270, 1215, 1040, 880, 830, 710, 688, 675. EIMS (probe, 70 eV) m/z (rel. int.): 324 [M]⁺ (1.2), 202 [M—benzoic acid]⁺ (12.1), 187 (5.6), 159 (9.4), 122 [benzoic acid]⁺ (60), 105 [benzoate]⁺ (100).

Fervanol *p*-hydroxybenzoate (2). Gum (34 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm⁻¹: 3360, 2970, 2940, 2870, 1680, 1610, 1595, 1512, 1450, 1365, 1280, 1220, 1160, 1100, 880, 850, 830, 770, 700. EIMS (probe, 70 eV) m/z (rel. int.): 340 [M]⁺ (0.8), 202 [M—*p*-hydroxybenzoic acid]⁺ (9.4), 187 (4.9), 159 (8.8), 138 [*p*-hydroxybenzoic acid]⁺ (40), 121 [*p*-hydroxybenzoate]⁺ (100).

Fervanol vanillate (3). Gum (9 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm⁻¹: 3400, 3080, 2975, 2938, 2870, 1710, 1610(sh), 1600, 1515, 1460, 1450, 1428, 1283, 1220, 1105, 1030, 880, 830, 785, 762, 725. EIMS (probe, 70 eV) m/z (rel. int.): 370 [M]⁺ (4), 202 [M—vanillic acid]⁺ (13.9), 187 (11.6), 168 [vanillic acid]⁺ (18), 159 (15.8), 151 [vanillate]⁺ (100).

Kurubasch aldehyde benzoate (4). Gum (45 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm⁻¹: 3060, 2970, 2930, 2865, 2710, 1720, 1695, 1602, 1586, 1490, 1450, 1318, 1270, 1110, 1098, 1028, 950, 935, 800, 710, 688, 672. EIMS (probe, 70 eV) m/z (rel. int.): 340 [M]⁺ (6.9), 218 [M—benzoic acid]⁺ (14.8), 203 (13.6), 189 (8.6), 175 (10), 122 [benzoic acid]⁺ (16.6), 105 [benzoate]⁺ (100).

Oxidation of 4. Kurubasch aldehyde benzoate (4) (10 mg) was dissolved in Et₂O (3 ml), chromic acid soln (0.3 ml) [20] was added dropwise and the soln stirred at room temp. for 1 hr. The Et₂O layer was separated and washed with NaHCO₃ soln (1 %), dried over anhyd. Na₂SO₄ and evapd *in vacuo* to yield 7 mg gum. Spectral properties of the product were found to be identical with 7.

Kurubasch aldehyde vanillate (5). Gum (11 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm⁻¹: 3400, 3080, 2970, 2940, 2870, 2715, 1710(sh), 1692, 1610(sh), 1600, 1512, 1460, 1430, 1388, 1370, 1280, 1215, 1100, 1032, 958, 878, 785, 762, 725. EIMS (probe, 70 eV) m/z (rel. int.): 386 [M]⁺ (3.3), 218 [M—vanillic acid]⁺ (5.9), 203 (2.3), 189 (3.1), 175 (4), 168 [vanillic acid]⁺ (15.9), 151 [vanillate]⁺ (100).

Kurubaschic acid angelate (6). Gum (25 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm⁻¹: 3100(sh), 2970, 2940, 2880, 2670, 2520(sh), 1715(sh), 1695, 1640, 1510, 1458, 1385, 1368, 1355, 1270, 1230, 1150, 1040, 960, 938, 845, 795. EIMS (probe, 70 eV) m/z (rel. int.): 334 [M]⁺ (1.1), 234 [M—angelic acid]⁺ (13), 219 (15.9), 206 (10.1), 191 (13.2), 149 (67.8), 105 (97), 100 [angelic acid]⁺ (68.6), 83 [angelate]⁺ (100).

Kurubaschic acid benzoate (7). Gum (230 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm⁻¹: 3100(sh), 3065, 2970, 2940, 2875, 2660, 2520(sh), 1720, 1692, 1640, 1602, 1585, 1512, 1450, 1388, 1370, 1320, 1270, 1210, 1108, 1070, 1026, 1002, 952, 938, 880, 796, 710, 688, 675. EIMS (probe, 70 eV) m/z (rel. int.): 356 [M]⁺ (0.14), 234 [M—benzoic acid]⁺ (32), 219 (33.9), 206 (5.7), 191 (24.7), 149 (38.5), 122 [benzoic acid]⁺ (82.4), 105 [benzoate]⁺ (100). CIMS (methane, probe, 70 eV) m/z (rel. int.): 355 [M—1]⁺ (5.9), 339 [M+1—H₂O]⁺ (6.4), 251 [M—1—benzoate+H]⁺ (9.8), 235 [M+1—benzoic acid]⁺ (100), 233 [M—1—benzoic acid]⁺ (62.5), 217 (43.2), 205 (20.3), 189 (48), 123 [benzoic acid+H]⁺ (59.8), 105 [benzoate]⁺ (51.5).

1 β ,10 α -Epoxykurubaschic acid angelate (8). Gum (12 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm⁻¹: 3200(sh), 2975, 2930, 2870, 2660, 2520(sh), 1715, 1695, 1645, 1510, 1453, 1385, 1270, 1260, 1230, 1150, 1040, 962, 862, 780, 710. EIMS (probe, 70 eV) m/z (rel. int.): 350 [M]⁺ (0.1), 250 [M—angelic acid]⁺ (4.4), 235 (2.7), 232 (4.2), 205 (5.2), 191 (6), 189 (6.6), 165 (12.9), 151 (17.7), 149 (17.6), 121 (19.3), 105 (28.9), 100 [angelic acid]⁺ (18.4), 83 [angelate]⁺ (100).

1 β ,10 α -Epoxykurubaschic acid benzoate (9). Gum (14 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm⁻¹: 3200(sh), 3060, 2970, 2940, 2875, 2660, 2520(sh),

1720, 1693, 1642, 1602, 1585, 1490, 1450, 1385, 1370, 1320, 1270, 1110, 1070, 1025, 955, 770, 710, 685, 677. EIMS (probe, 70 eV) m/z (rel. int.): 372 [M]⁺ (0.12), 250 [M—benzoic acid]⁺ (2.5), 235 (2.1), 232 (3.9), 205 (2), 191 (3.7), 189 (4.3), 165 (6.6), 151 (5.1), 122 [benzoic acid]⁺ (82.2), 105 [benzoate]⁺ (100). CIMS (methane, probe, 70 eV) m/z (rel. int.): 373 [M+1]⁺ (1.1), 371 [M—1]⁺ (1.3), 251 [M+1—benzoic acid]⁺ (20.6), 233 [M+1—benzoic acid—H₂O]⁺ (100), 223 (6.7), 215 (16.2), 205 (39.3), 193 (8.7), 187 (19.8), 177 (11.4), 123 [benzoic acid+H]⁺ (18.2), 105 [benzoate]⁺ (25.2).

1 α ,10 β -Epoxykurubaschic acid benzoate (10). Gum (8.5 mg); IR $\nu_{\text{max}}^{\text{NaCl}}$ cm⁻¹: 3200(sh), 3070, 2970, 2940, 2870, 2660, 2530(sh), 1720, 1695, 1645, 1602, 1588, 1490(sh), 1465, 1450, 1388, 1370, 1270, 1178, 1110, 1070, 1050, 1028, 960, 865, 775, 710, 688, 678. EIMS (probe, 70 eV) m/z (rel. int.): 272 [M]⁺ (0.3), 250 [M—benzoic acid]⁺ (1.8), 235 (1.4), 191 (3.1), 189 (3.9), 165 (6.4), 151 (3.7), 122 [benzoic acid]⁺ (56), 105 [benzoate]⁺ (100).

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REFERENCES

- Miski, M., Ulubelen, A. and Mabry, T. J. (1983) *Phytochemistry* **22**, 2231.
- Miski, M., Ulubelen, A., Mabry, T. J., Watson, W. H., Vickovic, I. and Holub, M. (1984) *Tetrahedron* **40**, 5197.
- Miski, M. and Mabry, T. J. (1985) *Phytochemistry* **24**, 1735.
- Miski, M. and Mabry, T. J. (1986) *Phytochemistry* **25**, 1673.
- Saidkhodzhaev, A. I. (1979) *Khim. Prir. Soedin.* 137.
- Sagitdinova, G. V. and Saidkhodzhaev, A. I. (1977) *Khim. Prir. Soedin.* 790.
- Sagitdinova, G. V., Saidkhodzhaev, A. I. and Malikov, V. M. (1978) *Khim. Prir. Soedin.* 809.
- Sagitdinova, G. V., Saidkhodzhaev, A. I. and Malikov, V. M. (1979) *Khim. Prir. Soedin.* 864.
- Buzhanova, K., Saidkhodzhaev, A. I. and Malikov, V. M. (1978) *Khim. Prir. Soedin.* 407.
- Buzhanova, K., Saidkhodzhaev, A. I. and Malikov, V. M. (1978) *Khim. Prir. Soedin.* 576.
- Sagitdinova, G. V., Saidkhodzhaev, A. I. and Malikov, V. M. (1982) *Khim. Prir. Soedin.* 721.
- Golovina, L. A. and Saidkhodzhaev, A. I. (1977) *Khim. Prir. Soedin.* 796.
- Bohlmann, F., Zdero, C. and Grenz, M. (1974) *Chem. Ber.* **107**, 3928.
- Bohlmann, F., Zdero, C., King, R. M. and Robinson, H. (1982) *Phytochemistry* **21**, 147.
- El Dahmy, S., Jakupovic, J., Bohlmann, F. and Sarg, T. M. (1985) *Tetrahedron* **41**, 309.
- Itokawa, H., Matsumoto, H., Mihashi, S. and Iitaka, Y. (1983) *Chem. Letters* 1581.
- Itokawa, H., Matsumoto, H., Mihashi, S., Iitaka, Y., Kasuya, A. and Itai, A. (1985) *Chem. Pharm. Bull.* **33**, 2204.
- Hendrickson, J. B. (1959) *Tetrahedron* **7**, 82.
- Sasaki, S., Itagaki, Y., Moriyama, H., Nakanishi, K., Watanabe, E. and Aoyama, T. (1966) *Tetrahedron Letters* 1623.
- Brown, H. C., Garg, C. P. and Liu, K.-T. (1971) *J. Org. Chem.* **36**, 387.